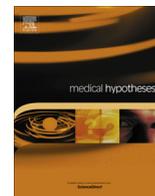


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Chronic and non-healing wounds: The story of vascular endothelial growth factor

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ABSTRACT

The pathophysiology of the chronicity and non-healing status of wounds remains unknown. This paper presents the following hypothesis: abnormal patterns of vascular endothelial growth factor receptors (VEGFRs) are the culprits of wound chronicity and non-healing. More specifically, for patients with poor circulation, the decreased VEGFR-2 level is the cause of poor wound healing; for patients with non-compromised circulation, for example, patients with concurrent chronic wounds and active autoimmune diseases, the increased VEGFR-1 level is related to the non-healing status of wounds.

The hypothesis is supported by the following facts. VEGFR-1 is the main contributor for inflammation and VEGFR-2 facilitates angiogenesis; soluble VEGFR-1 (sVEGFR-1) inactivates both VEGFR-1 and VEGFR-2. Patients with auto-immune disease have abnormally increased VEGFR-1 and decreased sVEGFR. Wounds in patients with active autoimmune diseases have poor response to electric stimulation which facilitates chronic wound healing in patients without active autoimmune diseases via increasing vascular endothelial growth factor (VEGF) secretion. Patients with chronic wounds (including diabetic foot ulcers and venous leg ulcers) but no active autoimmune diseases have decreased VEGFR-2 levels.

We thus believe that abnormal patterns of VEGFRs are the culprits of wound chronicity and non-healing. For wounds with compromised circulation, VEGFR-2 decrease contributes to its chronicity; whereas for wounds with non-compromised circulation, VEGFR-1 increase is the leading cause of the non-healing status of chronic wounds. Treatments and research in wound care should be tailored to target these changes based on circulation status of wounds. Complete elucidation of changes of VEGFRs in chronic and non-healing wounds will enhance our understandings in tissue healing and thus better our selection of appropriate treatments for chronic and non-healing wounds.

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Introduction

Approximately 6.5 million patients in the United States are being affected by chronic and non-healing wounds; the economic impact of chronic wound care is estimated to be US\$25 billion [1]. Although new and effective treatments, such as new collagen dressings and high voltage pulsed current therapy, are available for those suffering from open wounds, chronicity and delayed healing continue to impair lives and, in some cases, cause death [1]. Factors including poor circulation, ongoing pressure, systemic illnesses, age, and repeated trauma are known to contribute to the

chronicity or non-healing status of wounds [2]. Besides the limited understandings of detrimental effects of poor circulation to wound healing, the pathophysiology of the chronicity or non-healing status of wounds, especially in patients with sufficient blood supply to wound area, remains unknown. Pathophysiology of chronic wound healing is closely related to inflammation and angiogenesis. The vascular endothelial growth factors (VEGFs), a group of signal proteins, participate in critical rate-limiting steps of physiological vasculogenesis and angiogenesis [3]. Angiogenesis lays the foundation for granulation tissue development and the epithelialization process [3]. However, the role of VEGF in inflammation deserves better attention as abnormal high levels of VEGF contributes to the development of continuous inflammation [4,5]. Pro-inflammation effects of VEGF have been summarized as the following: (1) activates endothelial cells to secrete inflammatory

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cytokines and facilitates leukocyte adhesion and migration; (2) increases the production of tumor necrosis factor- α and interleukin-6 from mononucleocytes; (3) induces T cell transformation toward Th1 phenotype through increasing the production of interferon- γ and decreasing interleukin-10; (4) increases vascularity at the site of inflammation leading to more severe reactions [6]. Functions of VEGF depend on the activation of its receptors: activation of VEGF receptor-1 (VEGFR-1) leads to inflammation whereas activation of VEGF receptor-2 (VEGFR-2) causes angiogenesis; soluble VEGFR-1 (sVEGFR-1) sequesters VEGF, binds and inactivates membrane-bound VEGFR-1 and -2 receptors [4–6]. Electric stimulation therapy (E-stim) has been found to increase VEGF release and thus facilitates wound healing in various types of chronic wounds [3]. Research evidence from clinical trials indicates that E-stim improves wound healing in many patients, but not in others [3]. Based on our clinical experiences, wounds with non-optimal circulation usually respond better than wounds with simultaneous active autoimmune diseases but no compromise in circulation.

The hypothesis

Thus, this paper presents the following hypothesis: abnormal patterns of VEGFRs are the culprits of wound chronicity and non-healing. More specifically, for patients with poor circulation, the decreased VEGFR-2 level is the cause of poor wound healing; for patients without compromised circulation, for example, patients with concurrent chronic wounds and active autoimmune diseases, the increased VEGFR-1 level is the leading cause of the non-healing status of wounds. Thus, VEGF receptors may be used as biomarkers for predicting wound healing. In the following, we discussed the current understanding of VEGF and its receptors together with their participation in wound healing, the therapeutic mechanism of E-stim, as well as the abnormal patterns of VEGFRs in autoimmune diseases including systemic sclerosis (SS), rheumatoid arthritis (RA), and inflammatory bowel disease (IBD).

Current understandings of VEGF

Currently, the VEGF family is known to include five human VEGF isoforms (A, B, C, D, and placental growth factor-PLGF) which are produced by differential splicing of VEGF mRNA in endothelial cells, smooth muscle cells, platelets, neutrophils, keratinocytes, mast cells, monocytes/macrophages, and fibroblasts [7–10]. Effects of VEGF are mediated through binding of its subtypes to tyrosine kinase receptors (the VEGFRs) on the cell surface, causing them to dimerize and become activated through transphosphorylation [7–9]. Three main subtypes of VEGFR, numbered 1, 2 and 3, are available in the human body.

With tissue or skin injury, VEGFR-1 induces the migration of monocytes/macrophages, recruits hematopoietic stem cells, and induces the release of matrix metalloproteinases (MMPs) [11–13]. Additionally, VEGFR-1 is also the major contributor to inflammation [14–16] and a negative regulator for angiogenesis [17,18]. The negative effects of VEGFR-1 may be related to its inhibitory effects on VEGFR-2, even though the binding of PLGF with VEGFR-1 facilitates VEGFR-2 function [19–22]. As VEGFR-1 is a small protein that binds and sequesters VEGF-A to prevent it from binding to VEGFR-2 [23] and VEGFR-1 has at least 10 folds higher affinity in VEGF-A binding than VEGFR-2 [8,9], the negative modulation of VEGFR-2 by VEGFR-1 is likely caused by VEGFR-1 activation from VEGF-A.

Activation of VEGFR-2 is the major reason for mitogenic, angiogenic and permeability-enhancing effects, abnormal lymphangiogenesis, promoting cell adhesion and migration by VEGF-A [8,9].

It is worth noting that a large amount of research studies have found that the main contributor of angiogenesis is from the activation of VEGFR-2 with VEGF-A [6–9]. Stimulation of VEGFR-3 leads to the growth of lymphatic vessels without an influence on the blood vessel architecture [8]. Nonetheless, discussions on the function of VEGFR-3 are beyond the purposes of the present article. Thus, the function of VEGF-A depends on its activation of VEGFRs: activation of VEGFR-1 promotes inflammation; activation of VEGFR 2 mediates angiogenesis.

Inflammation and angiogenesis also involve the participation of other molecules, among which, MMPs, nitric oxide synthase, cyclooxygenase-2 (COX-2), hypoxia-inducible factor-1 α (HIF-1 α), and prostaglandin E2 have all been found to be involved in VEGF related pathophysiology [6,13]. Activation of VEGFR-1 with VEGF-A causes the induction and secretion of matrix metalloproteinase-9 (MMP-9) which further secretes VEGF-A [24–26]. VEGFR-2/VEGF-A activation leads to the release of nitric oxide and prostacyclin [27–32]. HIF-1 α targets genes of VEGF-A and VEGFR-2 and simulates their secretion [33,34]. Hypoxia also induces COX-2 expression which up-regulate HIF-1 α via increasing the formation of prostaglandin E2 [35]. Prostaglandin E2 is increased in chronic inflammatory sites and in tumor sites [36].

VEGF in chronic wound healing

Right after wounding, VEGF transcription and secretion are elevated and peak on day 3 and 7 in full thickness wounds and peak on day 2–3 on superficial wounds [37]. Although some researchers did not specify which specific subtype of VEGF is involved, the transcription and secretion of VEGF-A is more likely as the findings coincide with vascularity and granulation tissue formation and the majority of VEGF research studies in the early phase usually involve VEGF-A (VEGF165). In chronic wounds, the release of VEGF and other factors including platelet derived growth factor (PDGF), and fibroblast growth factor (FGF) is delayed or inhibited [38].

Research studies found that patients with diabetic foot ulcers have decreased VEGF and VEGFR-2 expression in tissues. VEGF mRNA levels are declined in wounds of diabetic mice as compared with normal mice [39]. Rivard et al. [40] reported that the reduced expression of VEGF in non-obese diabetic mice causes decreased angiogenesis and increasing VEGF expression could restore angiogenesis. Krishnan et al. [41] reported that epidermal and dermal blood vessel VEGFR-2 is decreased in patients with diabetic foot ulcer. However, angiopoietin-1 gene transfer improves impaired wound healing in genetically diabetic mice with augmented VEGFR-2 but without increasing VEGF expression [42].

Patients with venous insufficiency tend to have abnormal high plasma and tissue VEGF levels [43,44]. Lauer et al. [45] did biopsy on venous ulcers and found similarly increased VEGF expression in the wound epidermis area of chronic venous ulcer. Additionally, the expression of VEGFR-1 and 2 were found in papillary dermal microvessels near ulcer edge but not in capillaries distant to the ulcer edge [45]. Although increased expression of VEGF protein was detected in the epidermis, the intensity of this staining was weak in wound fluid compared with epidermis [45]. Simultaneously, sVEGFR-1 which sequesters VEGF, binds and inactivates membrane-bound VEGFR-1 and -2 receptors was reported high in wound fluid of chronic venous ulcer [46]. Although quantitative changes of tissue or plasma and tissue level VEGFR-1 and sVEGFR-1 remains unknown in venous leg ulcers, patients with non-healing venous leg ulcer are likely to have a low tissue/plasma VEGFR-2 level as sVEGFR-1 inactivates VEGFR-2.

Interestingly, researchers found that VEGF gene polymorphism is related to diabetic foot ulcer and the lower frequency of A allele in patients with diabetic foot ulcer is conferring a protective effect

[47]. Topical single-dose VEGF has no effect on wound healing [48]. Therefore, although VEGF function is essential for optimal wound angiogenesis, it may not be required for wound closure [49]. Consequently, we further believe that as compared to VEGF, VEGF receptors are more important for wound healing. The addition of VEGF apparently did not correct the deficiency of VEGFR-2 and thus likely rendered the introduction of VEGF to wound tissue non-relevant for wound healing in the previous studies [47–49].

Insights from E-stim in wound healing

Wounds in skin creates a direct current flow at the wound of at least 40 mV mm⁻¹ in cornea and up to 200 mV mm⁻¹ in guinea pig skin [50]. This electric field persists until re-epithelialization is complete [50]. During normal wound healing process, this endogenous electric field guides epithelial cell migration and nerve sprouting directly towards the wound edge [50]. Wound healing becomes compromised when the electric field is disrupted [50]. Wound-edge-directed nerve sprouting and angiogenesis are guided by the wound-induced electric field [50]. E-stim directly stimulate VEGF production by endothelial cells [50,51] and osteoblasts [52]. In muscle and skin of full-thickness wounds in animals, Asadi et al. [53] also reported that sensory and motor E-stim increased VEGF secretion. These effects of E-stim were also confirmed by the increase in serum and plasma VEGF in patients with dystrophic ulcers treated with E-stim [54].

To understand the roles of VEGFRs in E-stim for wound healing, Bai et al. [55] performed a study on human endothelial cells and found that E-stim upregulates VEGF-A and IL-8 secretion in human endothelial cells, and inhibition of VEGF receptor (VEGFR-1 or VEGFR-2) signaling significantly decreased VEGF production and completely abolished IL-8 production [55]. Electrical stimulation directly induces pre-angiogenic responses in vascular endothelial cells by signaling through VEGFRs [50]. Specifically, inhibition of VEGFR-2 completely abolished the E-stim-induced directional migration of the progenitor endothelial cell, thus researchers conclude that E-stim guides endothelial progenitor cell migration through VEGF receptor-2 signaling [56].

Insights learned from autoimmune diseases

In clinical practice, we observed that patients with concomitant active autoimmune disease and chronic wounds lacked favorable response to E-stim. To better elucidate the understandings of VEGFRs in wound healing, we thus reviewed changes of VEGF and its receptors in autoimmune diseases. VEGF-A which is the most widely studied VEGF isoform has been found to be significantly increased in patients with autoimmune disease, including Reynaud's phenomena in SS [4,57], uncontrolled RA (in both synovial fluid and serum) [58–61], and IBD (Crohn's disease and ulcerative colitis) [62]. Correspondingly, decrease or blockade in VEGF-A correlates with improved or decreased signs and symptoms in these disorders [61,62]. Recently, researchers found that genetic variation in the VEGF-A gene is associated with serum VEGF-A levels and correlates with the severity in RA symptoms in patients with RA [63–65]. Increase in serum VEGFR-1 has been reported in patients with RA, and correspondingly, specific blocking of VEGFR-1 has been found effective in decreasing inflammation in animal models with RA [66–68]. Increased expression of PLGF and VEGFR-1 was found in patients with RA indicating roles in rheumatoid inflammation by triggering production of proinflammatory cytokines while PLGF facilitates angiogenesis [68].

Although increased VEGF-A and VEGFR-2 levels have been reported in both patients with IBD and colitic mice, researchers

did not find increase in VEGFR-1 in intestine biopsies of patients with IBD [63]. Similar situations are true for patients with SS; no increase of VEGFR-1 was found in skin biopsy in patients with SS [69]. But, sVEGFR-1 secretions were reported to be low in tissue biopsies and inversely correlates with disease severity in patients with IBD and SS [70,71]. The artificial introduction or increase of sVEGFR-1 has been found to suppress angiogenesis and minimize inflammation damage in RA [72,73], IBD [62], and SS [71]. Interestingly, serum VEGFR-1 level was reported to be high in patients with Crohn's disease [72]. Increased serum VEGFR-1 in patients [72] seems contradictory to lack of increase in tissue level VEGFR-1 in animal studies of autoimmune diseases [68,69], but for wound healing, we tend to believe that serum level VEGFR-1 is more likely to be increased in patients with IBD and SS which is also supported with the decreased sVEGFR levels. As a result of the decreased sVEGFR-1 level in IBD and SS, these patients will likely have excessive functions of VEGFR-1 which is characterized by relentless inflammation in these patients.

To complete the loop related to the functions of VEGF and VEGFR-1 and -2, we also briefly reviewed a type of closely related molecule called vasohibin and other relevant molecules. The vasohibin family also participates in the VEGF system. Recent studies found that vasohibin-1 (VASH1) can be induced by both VEGF and FGF-2 [73]. Vasohibin-2 (VASH2) is not responsive to growth factors and cytokines and thus seems to be not inducible but constitutive [73]. Through studies on gene knockout mice, researchers identified that VASH1 inhibits angiogenesis at the termination zone and VASH2 stimulate angiogenesis in the sprout front [73]. VASH1 is up-regulated by VEGF in the retina and suppresses VEGFR-2 and retinal neovascularization [74]. Transient overexpression of VASH1 in human umbilical vein endothelial cells down-regulates VEGFR-1 and sVEGFR-1 on endothelial cells [75]. Expression of VASH1 in endothelial cells has been identified in various cancers, atherosclerotic lesions, age-dependent macular degeneration, diabetic retinopathy, RA and arterial re-endothelialization after denudation [73]. In RA, significant positive correlation exists between the expression of VASH1 and histological inflammation score ($p = 0.002$, $r = 0.842$) [76]. However, studies regarding the relationship between VASH1 and IBD or SS are lacking.

Discussion

Patients with RA tend to have simultaneous increase in VEGF-A, PLGF, VEGFR-1, and VASH1; patients with IBD and SS tend to have simultaneous increase in VEGF-A, VEGFR-1, VEGFR-2, and decreased sVEGFR. Characteristically, patients with RA, IBD, and SS all have abnormal VEGF and VEGF receptors (Table 1). VEGF-A binds VEGFR-1 leading to inflammation; VEGF-A or PLGF binds to VEGFR-2 leading to angiogenesis. VEGFR-1 inhibits VEGFR-2 (angiogenesis); sVEGFR-1 inhibits both VEGFR-1 and 2 (inflammation and angiogenesis); VASH1 inhibits sVEGFR-1 and VEGFR-2 (proinflammatory and inhibiting angiogenesis). As stated previously, VEGFR-2/VEGF-A activation leads to the release of nitric oxide and prostacyclin [28–33]. Nitric oxide generation was significantly improved with E-stim [54]. Antagonism of NO also inhibited VEGF production [77]. VEGFR-1/VEGF-A activation is involved in the induction of MMPs [14,26]. VEGFR-2/VEGF-A activation is involved in angiogenesis [6–9]. (Table 1 and Fig. 1.)

Wound healing process is closely related to angiogenesis (VEGFR-2) and inflammation (VEGFR-1). Chronic wounds can be divided into wounds with and without compromised circulation. Patients with diabetes, venous leg ulcers, and arterial wounds usually have compromised circulation. Patients with diabetic foot ulcers have decreased VEGF and VEGFR-2 expression in tissues;

Table 1
A summary of VEGF and its receptors.

RA (serum)	IBD & SS (serum)	Diabetic foot ulcer	Venous insufficiency/ulcer	Electric therapy
↑ VEGF	↑ VEGF	↓ VEGF in tissue	↑ VEGF in plasma and tissue of venous insufficiency	↑ VEGF
↑ VEGFR1	↑ VEGFR1, ↑ VEGFR-2	↓ VEGFR-2 in tissue	↑ sVEGFR-1 in non-healing venous wound fluid	
↑ PLGF	↓ sVEGFR1			
↑ VASH1				

Note: RA = rheumatoid arthritis; IBD = inflammatory bowel disease; SS = systemic sclerosis; VEGF = vascular endothelial growth factor-A; PGs = prostaglandins; NO = nitric oxide; sVEGFR1 = soluble vascular endothelial growth factor receptor 1; VASH1 = vasohibin 1; VEGFR1 and 2 = vascular endothelial growth factor receptor 1 and 2; PLGF = placental growth factor.

patients with venous insufficiency have increased VEGF in both plasma and tissue; patients with non-healing venous leg ulcers have high sVEGFR in the wound fluid which indicates possible low levels of VEGFR-2 in plasma and tissue. Consequently, for wounds with compromised circulation, the low level of VEGFR-2 is likely the cause for poor wound healing. Patients with concomitant chronic skin wounds and autoimmune disease usually have non-compromised circulation of the periwound tissue. Nonetheless, these patients also have abnormal patterns of VEGF receptors, more specifically, high VEGFR-1; non-healing skin wounds on patients with active autoimmune diseases are mainly characterized by inflammation. High VEGFR-1 level is thus likely responsible for the non-healing status of wounds without circulation compromise.

We thus believe that abnormal patterns of VEGFRs are the culprits of wound chronicity. Excessive inflammation impedes wound healing while predominant angiogenesis facilitates wound healing. VEGFR-1 is the vital molecule participating in the inflammation process whereas VEGFR-2 serves as the key factor in facilitating neovascularization. The normal physiological wound healing process requires balance between inflammation and angiogenesis. When a chronic wound is characterized by excessive inflammation which involves signs and symptoms of the heat (*calor*), pain (*dolor*), redness (*rubor*), and swelling (*tumor*), E-stim may not be beneficial but rather detrimental for wound healing. The rationales may be that increased VEGF release by E-stim will enhance the coupling of VEGF/VEGFR-1 aggravating inflammation which is common in patients with increased VEGFR-1. These patients, as exemplified by patients with concomitant autoimmune disease and chronic wounds, are likely to have relatively good circulation

of the periwound tissue. To the contrary, for patients with compromised circulation who usually do not show active inflammation, they are likely to have decreased VEGFR-2. These wounds are the majority of chronic wounds typically seen in clinical practices; they include arterial wounds, diabetic ulcers, venous leg wounds, and pressure ulcers. Thus, enhancement of angiogenesis and circulation is warranted to regain the physiological balance for wound healing via regulating VEGF and its receptors. As a beneficial tool to increase VEGF and activate VEGFR-2, E-stim will likely to facilitate wound healing in these patients.

To heal chronic wounds characterized by apparent inflammation with non-compromised circulation, therapies disrupting the VEGF-A/VEGFR-1 binding (proinflammation), more importantly decreasing VEGFR-1 activation, will be necessary. VEGFR-1 activation which is characterized by inflammation could be possibly decreased via the use of steroids. Although steroid use in chronic wound healing is a controversial topic, we believe the use of steroids in chronic and non-healing wounds with apparent inflammation are necessary and beneficial. Interestingly, this was supported by the study by Bosanquet et al. [78] in which researchers found that chronic wounds displaying abnormal inflammation could be successfully treated with topical steroids. Consequently, for the treatment of chronic and non-healing wounds with good periwound circulation, clinicians may facilitate wound healing through the following: (1) modify VEGF/VEGFR patterns; (2) modulate sVEGFR-1; (3) block inflammation with immunosuppressants including steroids.

To test these hypotheses, researchers investigating the pathophysiology of wound chronicity and non-healing should begin with classification of these wounds into two categories: wounds with and without compromised circulation. To explore the pathophysiology of the chronicity or non-healing status of wounds, studies should be performed to quantitatively identify VEGFRs in non-healing wounds as well as during the wound healing processes. Specifically, for wounds with non-compromised circulation, quantitative studies should be performed to identify changes of VEGFR-1; for chronic wounds with compromised circulation, emphasis should be the investigation of VEGFR-2. Complete elucidation of changes of VEGFRs in chronic and non-healing wounds will enhance our understandings in tissue healing and thus better our selection of appropriate treatments for chronic and non-healing wounds.

Conflicts of interest

The authors declare that no competing financial interests exist and this study did not receive any funding.

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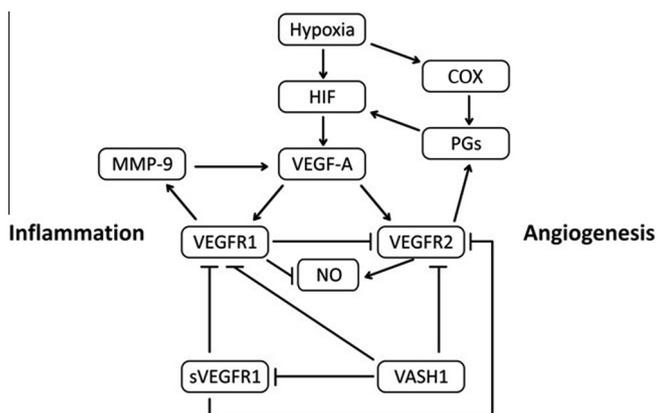


Fig. 1. Relationship among VEGF and related molecules in inflammation and angiogenesis. Note: VEGF-A = vascular endothelial growth factor-A; PGs = prostaglandins; NO = nitric oxide; sVEGFR1 = soluble vascular endothelial growth factor receptor 1; VASH1 = vasohibin1; VEGFR1 and 2 = vascular endothelial growth factor receptor 1 and 2. HIF = hypoxia induced factor; MMP = matrix metalloproteinases; COX = cyclooxygenase; ← represents activation; ⊥ represents inhibition.

References

- [1] Sen CK, Gordillo GM, Roy S, et al. Human skin wounds: a major and snowballing threat to public health and the economy. *Wound Repair Regen* 2009;17(6):763–71.
- [2] Guo S, Dipietro LA. Factors affecting wound healing. *J Dent Res* 2010;89(3):219–29.
- [3] Thakral G, Lafontaine J, Najafi B, et al. Electrical stimulation to accelerate wound healing. *Diabet Foot Ankle* 2013;4.
- [4] Manetti M, Guiducci S, Romano E, et al. Overexpression of VEGF165b, an inhibitory splice variant of vascular endothelial growth factor, leads to insufficient angiogenesis in patients with systemic sclerosis. *Circ Res* 2011;109:e14–26.
- [5] Yoo SA, Kwok SK, Kim WU. Proinflammatory role of vascular endothelial growth factor in the pathogenesis of rheumatoid arthritis: prospects for therapeutic intervention. *Mediators Inflamm* 2008;2008:129873.
- [6] Angelo LS, Kurzrock R. Vascular endothelial growth factor and its relationship to inflammatory mediators. *Clin Cancer Res* 2007;13(10):2825–30.
- [7] Olsson AK, Dimberg A, Kreuger J, et al. VEGF receptor signalling – in control of vascular function. *Nat Rev Mol Cell Biol* 2006;7(5):359–71.
- [8] Takahashi H, Shibuya M. The vascular endothelial growth factor (VEGF)/VEGF receptor system and its role under physiological and pathological conditions. *Clin Sci (Lond)* 2005;109(3):227–41.
- [9] Shibuya M. Vascular Endothelial Growth Factor (VEGF) and Its Receptor (VEGFR) Signaling in Angiogenesis: A Crucial Target for Anti- and Pro-Angiogenic Therapies. *Genes Cancer* 2011;2(12):1097–105.
- [10] Poesen K, Lambrechts D, Van Damme P, et al. Novel role for vascular endothelial growth factor (VEGF) receptor-1 and its ligand VEGF-B in motor neuron degeneration. *J Neurosci* 2008;28(42):10451–9.
- [11] Sawano A, Iwai S, Sakurai Y, et al. Flt-1, vascular endothelial growth factor receptor 1, is a novel cell surface marker for the lineage of monocyte-macrophages in humans. *Blood* 2001;97:785–91.
- [12] Hattori K, Heissig B, Wu Y, et al. Placental growth factor reconstitutes hematopoiesis by recruiting VEGFR1+ stem cells from bone-marrow microenvironment. *Nat Med* 2002;8:841–9.
- [13] Hiratsuka S, Nakamura K, Iwai S, et al. MMP9 induction by vascular endothelial growth factor receptor-1 is involved in lung specific metastasis. *Cancer Cell* 2002;2:289–300.
- [14] Mould AW, Tonks ID, Cahill MM, et al. Vegfb gene knockout mice display reduced pathology and synovial angiogenesis in both antigen-induced and collagen-induced models of arthritis. *Arthritis Rheum* 2003;48(9):2660–9.
- [15] Oura H, Bertoncini J, Velasco P, et al. A critical role of placental growth factor in the induction of inflammation and edema formation. *Blood* 2003;101(2):560–7.
- [16] Luttun A, Tjwa M, Moons L, et al. Revascularization of ischemic tissues by PlGF treatment, and inhibition of tumor angiogenesis, arthritis and atherosclerosis by anti-Flt1. *Nat Med* 2002;8(8):831–40.
- [17] Fong GH, Rossant J, Gertsentein M, Breitman ML. Role of the Flt-1 receptor tyrosine kinase in regulating the assembly of vascular endothelium. *Nature* 1995;376:66–70.
- [18] Hiratsuka S, Minowa O, Kuno J, Noda T, Shibuya M. Flt-1 lacking the tyrosine kinase domain is sufficient for normal development and angiogenesis in mice. *Proc Natl Acad Sci U S A* 1998;95:9349–54.
- [19] Zeng H, Dvorak HF, Mukhopadhyay D. Vascular permeability factor (VPF)/vascular endothelial growth factor (VEGF) receptor-1 down-modulates VPF/VEGF receptor-2-mediated endothelial cell proliferation, but not migration, through phosphatidylinositol 3-kinase-dependent pathways. *J Biol Chem* 2001;276(29):26969–79.
- [20] Zeng H, Sanyal S, Mukhopadhyay D. Tyrosine residues 951 and 1059 of vascular endothelial growth factor receptor-2 (KDR) are essential for vascular permeability factor/vascular endothelial growth factor-induced endothelium migration and proliferation, respectively. *J Biol Chem* 2001;276(35):32714–9.
- [21] Rahimi N, Dayanir V, Lashkari K. Receptor chimeras indicate that the vascular endothelial growth factor receptor-1 (VEGFR-1) modulates mitogenic activity of VEGFR-2 in endothelial cells. *J Biol Chem* 2000;275(22):16986–92.
- [22] Roberts DM, Kearney JB, Johnson JH, et al. The vascular endothelial growth factor (VEGF) receptor Flt-1 (VEGFR-1) modulates Flk-1 (VEGFR-2) signaling during blood vessel formation. *Am J Pathol* 2004;164(5):1531–5.
- [23] Gerber HP, Hillan KJ, Ryan AM, et al. VEGF is required for growth and survival in neonatal mice. *Development*. 1999;126:1149–59.
- [24] Fiorelli A, Morgillo F, Fasano M, et al. The value of matrix metalloproteinase-9 and vascular endothelial growth factor receptor 1 pathway in diagnosing indeterminate pleural effusion. *Interact Cardiovasc Thorac Surg* 2013;16(3):263–9.
- [25] Hollborn M, Stathopoulos C, Steffen A, et al. Positive feedback regulation between MMP-9 and VEGF in human RPE cells. *Invest Ophthalmol Vis Sci* 2007;48(9):4360–7.
- [26] Bergers G, Brekken R, McMahon G, et al. Matrix metalloproteinase-9 triggers the angiogenic switch during carcinogenesis. *Nat Cell Biol* 2000;2(10):737–44.
- [27] Mehta V, Abi-Nader KN, Peebles DM, et al. Long-term increase in uterine blood flow is achieved by local overexpression of VEGF-A(165) in the uterine arteries of pregnant sheep. *Gene Ther* 2012;19(9):925–35.
- [28] David AL, Torondel B, Zachary I, et al. Local delivery of VEGF adenovirus to the uterine artery increases vasorelaxation and uterine blood flow in the pregnant sheep. *Gene Ther* 2008;15(19):1344–50.
- [29] Laitinen M, Zachary I, Breier G, Pakkanen T, Hakkinen T, Luoma J, et al. VEGF gene transfer reduces intimal thickening via increased production of nitric oxide in carotid arteries. *Hum Gene Ther* 1997;8:1737–44.
- [30] Wheeler-Jones C, Abu-Ghazaleh R, Cospedal R, Houlston RA, Martin JF, Zachary I. Vascular endothelial growth factor stimulates prostacyclin production and activation of cytosolic phospholipase A2 in endothelial cells via p42/p44 mitogen-activated protein kinase. *FEBS Lett* 1997;420:28–32.
- [31] Horowitz JR, Rivard A, van der Zee R, Hariawala M, Sheriff DD, Esakof DD, et al. Vascular endothelial growth factor/vascular permeability factor produces nitric oxide-dependent hypotension. Evidence for a maintenance role in quiescent adult endothelium. *Arterioscler Thromb Vasc Biol* 1997;17:2793–800.
- [32] Li B, Ogasawara AK, Yang R, Wei W, He GW, Zioncheck TF, et al. KDR (VEGF receptor 2) is the major mediator for the hypotensive effect of VEGF. *Hypertension* 2002;39:1095–100.
- [33] Kurihara T, Westenskow PD, Friedlander M. Hypoxia-inducible factor (HIF)/vascular endothelial growth factor (VEGF) signaling in the retina. *Adv Exp Med Biol* 2014;801:275–81.
- [34] van der Horst IW, Rajatapiti P, van der Voorn P, et al. Expression of hypoxia-inducible factors, regulators, and target genes in congenital diaphragmatic hernia patients. *Pediatr Dev Pathol* 2011;14(5):384–90.
- [35] Jung YJ, Isaacs JS, Lee S, Trepel J, Neckers L. IL-1 β -mediated up-regulation of HIF-1 α via an NF- κ B/COX-2 pathway identifies HIF-1 as a critical link between inflammation and oncogenesis. *FASEB J* 2003;14:2115–7.
- [36] Abdel-Majid RM, Marshall JS. Prostaglandin E2 induces degranulation-independent production of vascular endothelial growth factor by human mast cells. *J Immunol* 2004;172:1227–36.
- [37] Bao P, Kodra A, Tomic-Canic M, et al. The role of vascular endothelial growth factor in wound healing. *J Surg Res* 2009;153(2):347–58.
- [38] Shukla A, Dubey MP, Srivastava R, et al. Differential expression of proteins during healing of cutaneous wounds in experimental normal and chronic models. *Biochem Biophys Res Commun* 1998;244(2):434–9.
- [39] Frank S, Hübner G, Breier G, et al. Regulation of vascular endothelial growth factor expression in cultured keratinocytes. Implications for normal and impaired wound healing. *J Biol Chem* 1995;270(21):12607–13.
- [40] Rivard A, Silver M, Chen D, et al. Rescue of diabetes-related impairment of angiogenesis by intramuscular gene therapy with adeno-VEGF. *Am J Pathol* 1999;154(2):355–63.
- [41] Krishnan ST, Quattrini C, Jeziorska M, et al. Neurovascular factors in wound healing in the foot skin of type 2 diabetic subjects. *Diabetes Care* 2007 Dec;30(12):3058–62.
- [42] Bitto A, Minutoli L, Galeano MR, et al. Angiopoietin-1 gene transfer improves impaired wound healing in genetically diabetic mice without increasing VEGF expression. *Clin Sci (Lond)* 2008 Jun;114(12):707–18.
- [43] Shoab SS, Scurr JH, Coleridge-Smith PD. Increased plasma vascular endothelial growth factor among patients with chronic venous disease. *J Vasc Surg* 1998;28(3):535–40.
- [44] Peschen M, Grenz H, Brand-Saberi B, et al. Increased expression of platelet-derived growth factor receptor alpha and beta and vascular endothelial growth factor in the skin of patients with chronic venous insufficiency. *Arch Dermatol Res* 1998;290(6):291–7.
- [45] Lauer G, Sollberg S, Cole M, et al. Expression and proteolysis of vascular endothelial growth factor is increased in chronic wounds. *J Invest Dermatol* 2000;115(1):12–8.
- [46] Eming SA, Lauer G, Cole M, et al. Increased levels of the soluble variant of the vascular endothelial growth factor receptor VEGFR-1 are associated with a poor prognosis in wound healing. *J Invest Dermatol* 2004;123(4):799–802.
- [47] Amoli MM, Hasani-Ranjbar S, Roohipour N, et al. VEGF gene polymorphism association with diabetic foot ulcer. *Diabetes Res Clin Pract* 2011;93(2):215–9.
- [48] Bolukbasi N, Balcioglu HA, Ozkan BT, et al. Topical Single-dose Vascular Endothelial Growth Factor has No Effect on Soft Tissue Healing. *N Am J Med Sci* 2014 Oct;6(10):505–9.
- [49] Jacobi J, Tam BY, Sundram U, et al. Discordant effects of a soluble VEGF receptor on wound healing and angiogenesis. *Gene Ther* 2004;11(3):302–9.
- [50] Zhao M, Bai H, Wang E, et al. Electrical stimulation directly induces pre-angiogenic responses in vascular endothelial cells by signaling through VEGF receptors. *J Cell Sci* 2004;117(Pt 3):397–405.
- [51] Bai H, Forrester JV, Zhao M. DC electric therapy upregulates angiogenic factors in endothelial cells through activation of VEGF receptors. *Cytokine* 2011 Jul;55(1):110–5.
- [52] Kim IS, Song JK, Zhang YL, et al. Biphasic electric current stimulates proliferation and induces VEGF production in osteoblasts. *Biochim Biophys Acta* 2006;1763(9):907–16.
- [53] Asadi MR, Torkaman G, Hedayati M. Effect of sensory and motor electrical stimulation in vascular endothelial growth factor expression of muscle and skin in full-thickness wound. *JRRD* 2011;48:195.
- [54] Ferroni P, Roselli M, Guadagni F, et al. Biological effects of a software-controlled voltage pulse generator (PhyBack PBK-2C) on the release of vascular endothelial growth factor (VEGF). *In Vivo* 2005;19(6):949–58.
- [55] Bai H, Forrester JV, Zhao M. C electric therapy upregulates angiogenic factors in endothelial cells through activation of VEGF receptors. *Cytokine* 2011;55(1):110–5.
- [56] Zhao Z, Qin L, Reid B, Pu J, Hara T, Zhao M. Directing migration of endothelial progenitor cells with applied DC electric fields. *Stem Cell Res* 2012 Jan;8(1):38–48.

- [57] Maurer B, Distler A, Suliman YA, et al. Vascular endothelial growth factor aggravates fibrosis and vasculopathy in experimental models of systemic sclerosis. *Ann Rheum Dis* 2014;73(10):1880–7.
- [58] Nagashima M, Yoshino S, Aono H, et al. Inhibitory effects of anti-rheumatic drugs on vascular endothelial growth factor in cultured rheumatoid synovial cells. *Clin Exp Immunol* 1999 May;116(2):360–5.
- [59] Vordenbäumen S, Sewerin P, Lögters T, et al. Inflammation and vascularisation markers of arthroscopically-guided finger joint synovial biopsies reflect global disease activity in rheumatoid arthritis. *Clin Exp Rheumatol* 2014;32(1):117–20.
- [60] Lahoti TS, Hughes JM, Kusnadi A, et al. Aryl hydrocarbon receptor antagonism attenuates growth factor expression, proliferation, and migration in fibroblast-like synoviocytes from patients with rheumatoid arthritis. *J Pharmacol Exp Ther* 2014;348(2):236–45.
- [61] Taylor PC. VEGF and imaging of vessels in rheumatoid arthritis. *Arthritis Res* 2002;4(Suppl 3):S99–107.
- [62] Scaldaferrì F, Vetrano S, Sans M, et al. VEGF-A links angiogenesis and inflammation in inflammatory bowel disease pathogenesis. *Gastroenterology* 2009;136(2):585–595.e5.
- [63] Chen Y, Dawes PT, Matthey DL. Polymorphism in the vascular endothelial growth factor A (VEGFA) gene is associated with serum VEGF – a level and disease activity in rheumatoid arthritis: differential effect of cigarette smoking. *Cytokine* 2012;58(3):390–7.
- [64] Zhang Y, Qiu H, Zhang H, et al. Vascular endothelial growth factor A (VEGFA) polymorphisms in Chinese patients with rheumatoid arthritis. *Scand J Rheumatol* 2013;42(5):344–8.
- [65] Chen Y, Matthey DL. Age at onset of rheumatoid arthritis: association with polymorphisms in the vascular endothelial growth factor A(VEGFA) gene and an intergenic locus between matrix metalloproteinase (MMP) 1 and 3 genes. *Clin Exp Rheumatol* 2012;30(6):894–8.
- [66] De Bandt M, Ben Mahdi MH, Ollivier V, et al. Blockade of vascular endothelial growth factor receptor I (VEGF-RI), but not VEGF-RII, suppresses joint destruction in the K/BxN model of rheumatoid arthritis. *J Immunol* 2003;171(9):4853–9.
- [67] Murakami M, Iwai S, Hiratsuka S, et al. Signaling of vascular endothelial growth factor receptor-1 tyrosine kinase promotes rheumatoid arthritis through activation of monocytes/macrophages. *Blood* 2006;108(6):1849–56.
- [68] Yoo SA, Yoon HJ, Kim HS, et al. Role of placenta growth factor and its receptor flt-1 in rheumatoid inflammation: a link between angiogenesis and inflammation. *Arthritis Rheum* 2009;60(2):345–54.
- [69] Higashi-Kuwata N, Makino T, Inoue Y, et al. Expression pattern of VEGFR-1, -2, -3 and D2–40 protein in the skin of patients with systemic sclerosis. *Eur J Dermatol* 2011;21(4):490–4.
- [70] Wejman J, Pyzlak M, Szukiewicz D, et al. Thrombospondin and VEGF-R: is there a correlation in inflammatory bowel disease? *Mediators Inflamm* 2013;2013:908259.
- [71] Avouac J, Wipff J, Goldman O, et al. Angiogenesis in systemic sclerosis: Impaired expression of vascular endothelial growth factor receptor 1 in endothelial progenitor-derived cells under hypoxic conditions. *Arthritis Rheum* 2008;58(11):3550–61.
- [72] Dueñas Pousa I, Maté Jiménez J, Salcedo Mora X, et al. Analysis of soluble angiogenic factors in Crohn's disease: a preliminary study. *Gastroenterol Hepatol* 2007;30(9):518–24.
- [73] Sato Y. The vasohibin family: a novel family for angiogenesis regulation. *J Biochem* 2013;153(1):5–11.
- [74] Shen J, Yang X, Xiao WH, et al. Vasohibin is up-regulated by VEGF in the retina and suppresses VEGF receptor 2 and retinal neovascularization. *FASEB J* 2006;20(6):723–5.
- [75] Miyashita H, Suzuki H, Ohkuchi A, et al. Mutual balance between vasohibin-1 and soluble VEGFR-1 in endothelial cells. *Pharmaceuticals* 2011;4:1551–77.
- [76] Miyake K, Nishida K, Kadota Y, et al. Inflammatory cytokine-induced expression of vasohibin-1 by rheumatoid synovial fibroblasts. *Acta Med Okayama* 2009;63(6):349–58.
- [77] Kilani MM, Mohammed KA, Nasreen N, et al. RSV causes HIF-1 α stabilization via NO release in primary bronchial epithelial cells. *Inflammation* 2004 Oct;28(5):245–51.
- [78] Bosanquet DC, Rangaraj A, Richards AJ, et al. Topical steroids for chronic wounds displaying abnormal inflammation. *Ann R Coll Surg Engl* 2013;95(4):291–6.